

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION  
International Bureau



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 7 : <b>C07K 14/00</b>		<b>A2</b>	(11) International Publication Number: <b>WO 00/69900</b>
			(43) International Publication Date: 23 November 2000 (23.11.00)
(21) International Application Number: <b>PCT/US00/13576</b>		(74) Agents: WARD, Michael, R. et al.; Limbach & Limbach L.L.P., 2001 Ferry Building, San Francisco, CA 94111-4207 (US).	
(22) International Filing Date: 17 May 2000 (17.05.00)			
(30) Priority Data: 60/134,406 17 May 1999 (17.05.99) US 60/153,406 10 September 1999 (10.09.99) US 60/159,783 15 October 1999 (15.10.99) US		(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).	
(71) Applicant (for all designated States except US): CONJUCHEM, INC. [CA/CA]; Suite 3950, Third Floor, 225 President Kennedy Avenue West, Montreal, Quebec H2X 3Y8 (CA).		<b>Published</b> <i>Without international search report and to be republished upon receipt of that report.</i>	
(72) Inventors; and (75) Inventors/Applicants (for US only): BRIDON, Dominique, P. [FR/CA]; 243 chemin Cote Ste-Catherine, Outremont, Quebec H2V 2B2 (CA). EZRIN, Alan, M. [US/US]; 110 Quintas Lane, Moraga, CA 94556 (US). MILNER, Peter, G. [GB/US]; 14690 Manuella Road, Los Altos Hills, CA 94022 (US). HOLMES, Darren, L. [US/CA]; 3450 Drummond Street, Montreal, Quebec H3G 1T3 (CA). THIBAudeau, Karen [FR/CA]; 4700 Bonavista Street, #407, Montreal, Quebec H3W 2L5 (CA).			
(54) Title: PROTECTION OF ENDOGENOUS THERAPEUTIC PEPTIDES FROM PEPTIDASE ACTIVITY THROUGH CONJUGATION TO BLOOD COMPONENTS			
(57) Abstract <p>A method for protecting a peptide from peptidase activity <i>in vivo</i>, the peptide being composed of between 2 and 50 amino acids and having a C-terminus and an N-terminus and a C-terminus amino acid and an N-terminus amino acid is described. In the first step of the method, the peptide is modified by attaching a reactive group to the C-terminus amino acid, to the N-terminus amino acid, or to an amino acid located between the N-terminus and the C-terminus, such that the modified peptide is capable of forming a covalent bond <i>in vivo</i> with a reactive functionality on a blood component. In the next step, a covalent bond is formed between the reactive group and a reactive functionality on a blood component to form a peptide-blood component conjugate, thereby protecting said peptide from peptidase activity. The final step of the method involves the analyzing of the stability of the peptide-blood component conjugate to assess the protection of the peptide from peptidase activity.</p>			

Relevant pages only.

total pages 433

- 98 -

dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et<sub>2</sub>O (Step 4). The product is purified by preparative reverse phase HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H<sub>2</sub>O (A) and 0.045% TFA in CH<sub>3</sub>CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10  $\mu$  phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at  $\lambda$  214 and 254 nm to afford the desired DAC in >95% purity, as determined by RP-HPLC.

**Example 8 – Modification of Dyn A 1-13 at the  $\epsilon$ -Amino Group of the Added C-terminus Lysine Residue - Synthesis of Dyn A 1-13(N $\epsilon$ -MPA)-NH<sub>2</sub>**  
15 **Tyr-Gly-Gly-Phe-Leu-Arg-Arg-Ile-Arg-Pro-Lys-Leu-Lys-(N $\epsilon$ -MPA)-NH<sub>2</sub>**

Solid phase peptide synthesis of a modified Dyn A 1-13 on a 100  $\mu$ mole scale was performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids were sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Pro-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Gly-OH, Fmoc-Gly-OH, Fmoc-Tyr(tBu)-OH. They were dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (Step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh<sub>3</sub>)<sub>4</sub> dissolved in 5 mL of CHCl<sub>3</sub>:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl<sub>3</sub> (6 x 5 mL), 20% HOAc in DCM (6 x 5

- 99 -

mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is  
 5 cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et<sub>2</sub>O (Step 4). The product is purified by preparative reverse phase HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H<sub>2</sub>O (A) and 0.045% TFA in CH<sub>3</sub>CN (B)) over 180 min at  
 10 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired DAC in >95% purity, as determined by RP-HPLC.

The structure of this product is



15

**Example 9 - Modification of Dyn A 2-13 at the ε-Amino Group of the Added C-terminus Lysine Residue - Synthesis of Dyn A 2-13(Nε-MPA)-NH<sub>2</sub>**  
 20 Gly-Gly-Phe-Leu-Arg-Arg-Ile-Arg-Pro-Lys-Leu-Lys-(Nε-MPA)-NH<sub>2</sub>

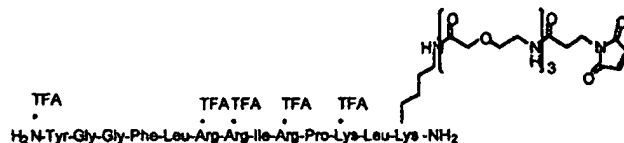
Using automated peptide synthesis, the following protected amino acids were sequentially added to Rink Amide MBHA resin: Fmoc-  
 25 Lys(Mtt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Pro-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Gly-OH, and Boc-Gly-OH. Manual synthesis was employed for the remaining steps: selective removal of the Mtt group and coupling of MPA using HBTU/HOBt/DIEA activation in

- 100 -

DMF. The target dynorphin analog was removed from the resin; the product was isolated by precipitation and purified by preparative HPLC to afford the desired product as a white solid upon lyophilization in a 35% yield. Anal. HPLC indicated product to be >95% pure with  $R_t =$   
 5 30.42 min. ESI-MS  $m/z$  for  $C_{73}H_{123}N_{25}O_{15}$  ( $MH^+$ ), calcd 1590.0, found  $MH^{3+}$  531.3.

**Example 10 - Modification of Dyn A 1-13 at the  $\epsilon$ -Amino Group of the Added C-terminus Lysine Residue - Synthesis of Dyn A 1-13(AEA<sub>3</sub>-MPA)-NH<sub>2</sub>**  
 10 **Tyr-Gly-Gly-Phe-Leu-Arg-Arg-Ile-Arg-Pro-Lys-Leu-Lys-(AEA<sub>3</sub>-MPA)-NH<sub>2</sub>**

Using automated peptide synthesis, the following protected amino acids were sequentially added to Rink Amide MBHA resin: Fmoc-  
 15 Lys(Mtt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Pro-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Gly-OH, Fmoc-Gly-OH, and Boc-Tyr(Boc)-OH. Manual synthesis was employed for the remaining steps: selective removal of the Mtt group, the coupling of three-Fmoc-AEA-OH groups (AEA = aminoethoxyacetic acid) with Fmoc removal in-between  
 20 each coupling, and MPA acid using HBTU/HOBt/DIEA activation in DMF. The target dynorphin analog was removed from the resin; the product was isolated by precipitation and purified by preparative HPLC to afford the desired product as a white solid upon lyophilization in a  
 25 29% yield. Anal. HPLC indicated product to be >95% pure with  $R_t =$  33.06 min. ESI-MS  $m/z$  for  $C_{94}H_{154}N_{29}O_{23}$  ( $MH^+$ ), calcd 2057.2, found  $MH^{4+}$  515.4,  $MH^{3+}$  686.9,  $MH^{2+}$  1029.7.



- 101 -

**Example 11 – Modification of Dyn A 2-13 at the  $\epsilon$ -Amino Group of the Added C-terminus Lysine Residue - Synthesis of Dyn A 2-13(AEA<sub>3</sub>-MPA)-NH<sub>2</sub>**  
**Gly-Gly-Phe-Leu-Arg-Arg-Ile-Arg-Pro-Lys-Leu-Lys-(AEA<sub>3</sub>-MPA)-NH<sub>2</sub>**

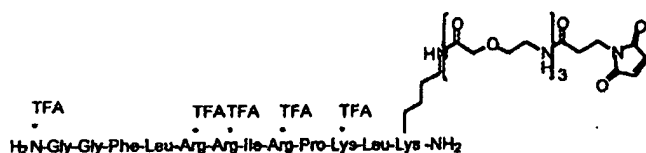
5

Using automated peptide synthesis, the following protected amino acids were sequentially added to Rink Amide MBHA resin: Fmoc-Lys(Mtt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Pro-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Gly-OH, and Fmoc-Gly-OH. Manual synthesis was employed for the remaining steps: selective removal of the Mtt group, the coupling of three-Fmoc-AEA-OH groups, with Fmoc removal in-between each coupling, and MPA using HBTU/HOBt/DIEA activation in DMF. The target dynorphin analog was removed from the resin; the product was isolated by precipitation and purified by preparative HPLC to afford the desired product as a white solid upon lyophilization in a 29% yield. Anal. HPLC indicated product to be >95% pure with  $R_t = 31.88$  min. ESI-MS  $m/z$  for C<sub>85</sub>H<sub>145</sub>N<sub>25</sub>O<sub>21</sub> (MH<sup>+</sup>), calcd 1894.3, found MH<sup>4+</sup> 474.6, MH<sup>3+</sup> 632.4, MH<sup>2+</sup> 948.10.

10

15

20



**Example 12 – Modification of Neuropeptide Y at the  $\epsilon$ -Amino Group of the Added C-terminus Lysine Residue**  
**Preparation of Tyr-Pro-Ser-Lys-Pro-Asp-Asn-Pro-Gly-Glu-Asp-Ala-Pro-Ala-Glu-Asp-Met-Ala-Arg-Tyr-Tyr-Ser-Ala-Leu-Lys-(N- $\epsilon$ -MPA)-NH<sub>2</sub>**

25

30

Solid phase peptide synthesis of a modified neuropeptide Y analog on a 100  $\mu$ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink

- 102 -

Amide MBHA. The following protected amino acids were sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Ser(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ala-OH, Fmoc-Met-OH, Fmoc-Asp(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ala-OH, Fmoc-Pro-OH, Fmoc-Ala-OH, Fmoc-Asp(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gly-OH, Fmoc-Pro-OH, Fmoc-Asn(Trt)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Pro-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Pro-OH, Fmoc-Tyr(tBu)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N,N*-tetramethyluronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (Step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh<sub>3</sub>)<sub>4</sub> dissolved in 5 mL of CHCl<sub>3</sub>:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl<sub>3</sub> (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et<sub>2</sub>O (Step 4). The product is purified by preparative reverse phase HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H<sub>2</sub>O (A) and 0.045% TFA in CH<sub>3</sub>CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenylhexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired DAC in >95% purity, as determined by RP-HPLC.

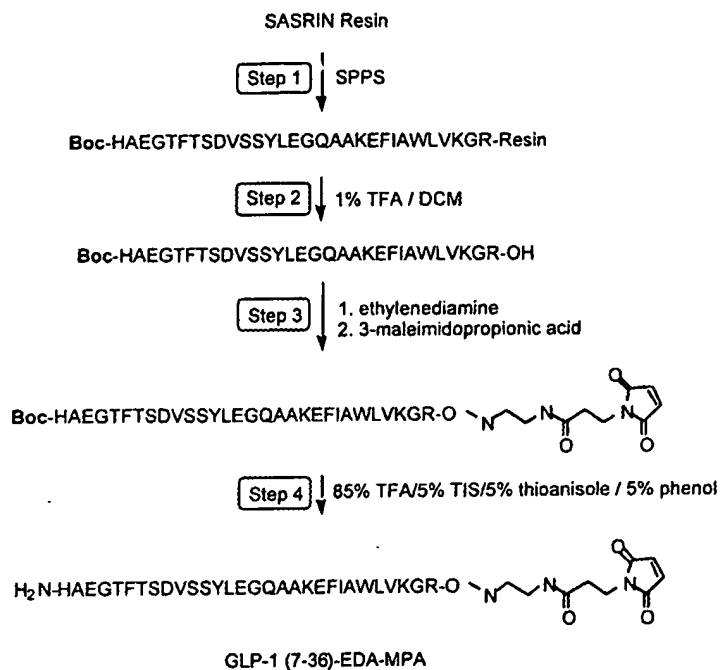
**Example 13 - Modification of GLP-1 (7-36) at the C-Terminus  
Arginine  
Preparation of GLP-1 (7-36)-EDA-MPA**

- 5 Solid phase peptide synthesis of a modified GLP-1 analog on a 100  $\mu$ mole scale is performed manually and on a Symphony Peptide Synthesizer SASRIN (super acid sensitive resin). The following protected amino acids are sequentially added to the resin: Fmoc-Arg(Pbf)-OH, Fmoc-Gly-OH, Fmoc-Lys(Boc)-OH, Fmoc-Val-OH, Fmoc-  
10 Leu-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ala-OH, Fmoc-Ile-OH, Fmoc-Phe-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Ala-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gly-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Leu-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-  
15 OH, Fmoc-Phe-OH, Fmoc-Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Ala-OH, Boc-His(Trt)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N,N*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA).  
20 Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (Step 1). The fully protected peptide is cleaved from the resin by treatment with 1% TFA / DCM (Step 2). Ethylenediamine and 3-maleimidopropionic acid are then sequentially added to the free C-  
25 terminus (Step 3). The protecting groups are then cleaved and the product isolated using 86% TFA/5% TIS/5% H<sub>2</sub>O/2% thioanisole and 2% phenol, followed by precipitation by dry-ice cold Et<sub>2</sub>O (Step 4). The product is purified by preparative reverse phase HPLC using a Varian (Rainin) preparative binary HPLC system using a Dynamax C<sub>18</sub>, 60Å, 8  
30  $\mu$ m, 21 mm x 25 cm column equipped with a Dynamax C<sub>18</sub>, 60Å, 8  $\mu$ m guard module, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at  $\lambda$  214 and 254 nm to afford the desired DAC in

- 104 -

>95% purity, as determined by RP-HPLC. These steps are illustrated in the schematic diagram below.

5



**Example 14 - Modification of Exendin-4 at the C-terminus Serine Preparation of Exendin-4 (1-39)-EDA-MPA**

10 Solid phase peptide synthesis of a modified Exendin-4 analog on a 100 μmole scale is performed manually and on a Symphony Peptide Synthesizer SASRIN (super acid sensitive resin). The following protected amino acids are sequentially added to the resin: Fmoc-

15 Ser(tBu)-OH, Fmoc-Pro-OH, Fmoc-Pro-OH, Fmoc-Pro-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Pro-OH, Fmoc-Gly-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Leu-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Ile-



- 105 -

OH, Fmoc-Phe-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Met-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(OtBu)-OH, Fmoc--

5 Ser(tBu)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Phe-OH, Fmoc-Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Gly-OH, Boc-His(Trt)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N, N, N, N*-tetramethyl-uronium hexafluorophosphate (HBTU) and

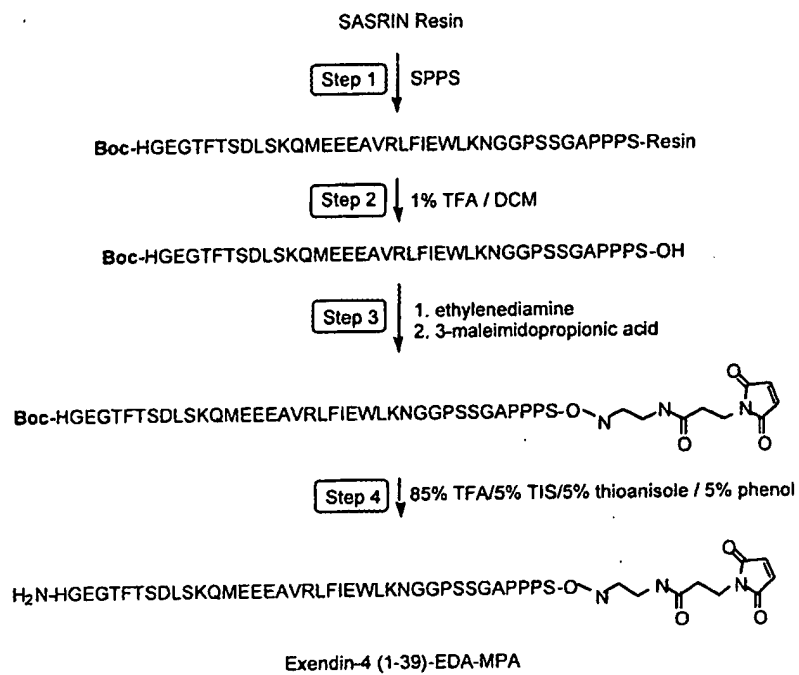
10 Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (Step 1). The fully protected peptide is cleaved from the resin by treatment with 1% TFA / DCM (Step 2). Ethylenediamine and 3-maleimidopropionic acid are then sequentially

15 added to the free C-terminus (Step 3). The protecting groups are then cleaved and the product isolated using 86% TFA/5% TIS/5% H<sub>2</sub>O/2% thioanisole and 2% phenol, followed by precipitation by dry-ice cold Et<sub>2</sub>O (Step 4). The product is purified by preparative reverse phase HPLC using a Varian (Rainin) preparative binary HPLC system using a

20 Dynamax C<sub>18</sub>, 60Å, 8 µm, 21 mm x 25 cm column equipped with a Dynamax C<sub>18</sub>, 60Å, 8 µm guard module, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired DAC in >95% purity, as determined by RP-HPLC.

25

- 106 -



5    **Example 15 – Modification of Secretin Peptide at the  $\epsilon$ -Amino Group of the Added C-terminus Lysine Residue**  
Preparation of His-Ser-Asp-Gly-Thr-Phe-Thr-Ser-Glu-Leu-Ser-Arg-Leu-Arg-Glu-Gly-Ala-Arg-Leu-Glu-Arg-Leu-Leu-Gln-Gly-Leu-Val-Lys-(N $\epsilon$ -MPA)-NH<sub>2</sub>

10            Solid phase peptide synthesis of a modified secretin peptide analog on a 100  $\mu$ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially  
15    added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Val-OH, Fmoc-Leu-OH, Fmoc-Gly-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-Glu(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH,  
20    Fmoc-Glu(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Phe-OH, Fmoc-Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-His(Boc)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium  
25    hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (Step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq  
30    of Pd(PPh<sub>3</sub>)<sub>4</sub> dissolved in 5 mL of CHCl<sub>3</sub>:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl<sub>3</sub> (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-  
35    dimethylformamide (DMF) and 3 times with isopropanol. The peptide is

cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et<sub>2</sub>O (Step 4). The product is purified by preparative reverse phase HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H<sub>2</sub>O (A) and 0.045% TFA in CH<sub>3</sub>CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10  $\mu$  phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at  $\lambda$  214 and 254 nm to afford the desired DAC in >95% purity, as determined by RP-HPLC.

**Example 16 – Modification of Kringle-5 at the  $\epsilon$ -Amino Group of the Added C-terminus Lysine Residue**  
**Preparation of NAc-Pro-Arg-Lys-Leu-Tyr-Asp-Tyr-Lys-(N $\epsilon$ -MPA)-NH<sub>2</sub>.2TFA**

Solid phase peptide synthesis of a modified Kringle-5 peptide on a 100  $\mu$ mole scale was performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Pro-OH. They were dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). At the end of the synthesis Acetic Anhydride was added to acetylate the N-terminal. The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh<sub>3</sub>)<sub>4</sub> dissolved in 5 mL of CHCl<sub>3</sub>:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl<sub>3</sub> (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3).